

Test Validation

- The mean of the **Negative Controls** must produce an optical density ≥ 0.300 and ≤ 2.000 .
- The mean of the **Positive Controls** must have an inhibition of $\geq 40\%$.

Interpreting the Results

- If a test sample produces $\geq 40\%$ inhibition, it is positive.
- If a test sample produces $< 40\%$ inhibition, it is negative.

Precautions

Kit components should be handled and disposed of as potentially hazardous. Do not eat, drink, or smoke where serum samples and kit reagents are handled. Do not pipette by mouth. Some reagents may be harmful if ingested. If ingested, seek medical attention. Do not use expired or contaminated reagents, or reagents from other kits or serials. Do not mix reagents from different serials of this same product.

Component B, Positive Control, contains sodium azide as a preservative.

Component C, Negative Control, contains sodium azide as a preservative.

Component D, 100X Primary Antibody, contains ProClin 300, methylisothiazolone, and bromonitrodioxane as preservatives.

Component E, 100X Secondary Antibody-Peroxidase Conjugate, contains ProClin 300, methylisothiazolone, bromonitrodioxane, and thimerosal as preservatives.

Component F, Antibody Diluting Buffer, contains ProClin 300, methylisothiazolone and bromonitrodioxane as preservatives.

Component G, Serum Diluting Buffer, contains sodium azide as a preservative.

Component J, Stop Solution, contains sodium fluoride.

USDA Veterinary License No. 332

Version 161101

BABESIA CABALLI ANTIBODY TEST KIT, cELISA

Assay instructions for catalog number 273-2

General Description

This competitive, enzyme-linked, immunosorbent assay (cELISA) detects antibodies to *Babesia caballi* in equine sera. Sample serum *B. caballi* antibody inhibits binding of primary monoclonal antibody. The binding of primary monoclonal antibody to the antigen-coated plate is detected with horseradish-peroxidase (HRP)-labeled secondary antibody. Binding of the HRP-labeled secondary antibody is detected by the addition of enzyme substrate and quantified by subsequent color product development. Strong color development indicates little or no inhibition of primary monoclonal antibody binding and therefore the absence of *B. caballi* antibody in sample sera. Weak color development due to inhibition of the primary monoclonal antibody binding to the antigen on the solid phase indicates the presence of *B. caballi* antibodies in sample sera.

Kit Contents

Component		
A	Antigen-Coated Plates	2 plates
B	Positive Control	2 ml
C	Negative Control	2 ml
D	100X Primary Antibody	0.3 ml
E	100X Secondary Antibody-Peroxidase Conjugate	0.3 ml
F	Antibody Diluting Buffer	60 ml
G	Serum Diluting Buffer	10.5 ml
H	10X Wash Solution Concentrate	120 ml
I	Substrate Solution	30 ml
J	Stop Solution	30 ml
Instructions for Use		

Materials Required But Not Included in the Test Kit

Single and multichannel adjustable-volume pipettors and disposable plastic tips, non-antigen-coated transfer plate(s), ELISA microplate absorbance spectrophotometer with 620, 630 or 650 nm filter, deionized or distilled water, paper towels, graduated cylinder, timer, multichannel pipettor reservoirs, wash bottle, manual multichannel washing device or automatic plate washer

Storage and Stability

Store all reagents at 2-7°C. **Do not freeze.** Reagents will remain stable until the expiration date when stored as instructed. **Do not use test kit past the expiration date.**

Preparation

- a. **Warm reagents:** Bring the serum samples, reagents and plate(s) to room temperature ($23 \pm 2^\circ\text{C}$) prior to starting the test.
- b. **Prepare controls and samples:** The Positive and Negative Controls (B & C) and test serum samples must be diluted 1/2 in Serum Diluting Buffer (G) for use in this test. It is recommended that these dilutions be made in a non-antigen-coated transfer plate. Add Serum Diluting Buffer (G) to the appropriate wells of a transfer plate according to your setup record. Add an equal volume of Positive Control (B), Negative Control (C), or sample to each well and cycle the pipettor at least three times to mix. Change tips between samples. The volume in the transfer plate must be in excess of 50 μl in order to transfer 50 μl from it. Load Positive Control (B) in duplicate and Negative Control (C) in triplicate, regardless of the number of serum samples to be tested. When whole plates are used, it is best to put the controls in wells on different areas of the plate. The controls must be loaded every time the assay is performed and on each plate if multiple plates are used.
- c. **Prepare plates:** Remove the plate(s) from the foil pouch(es) (A). If applicable: Return any unused strips to the pouch and securely seal it. Place strips to be used in the frame and number the top of each strip to maintain orientation. Always mark the strips in case they dislodge from the frame during washing.
- d. **Prepare primary antibody:** Prepare 1X Primary Antibody by diluting 1 part of the 100X Primary Antibody (D) with 99 parts of Antibody Diluting buffer (F). Example: For 96 wells, mix 60 μl of 100X Primary Antibody (D) with 5.940 ml of Antibody Diluting Buffer (F) to yield 6 ml of 1X Primary Antibody. Fifty microliters (50 μl) are needed per well. Allow extra quantity for reservoirs, tubing, pipetting, etc.
- e. **Prepare secondary antibody-peroxidase conjugate:** Prepare 1X Secondary Antibody-Peroxidase Conjugate by diluting 1 part of the 100X Secondary Antibody-Peroxidase Conjugate (E) with 99 parts of Antibody Diluting buffer (F). Example: For 96 wells, mix 60 μl of 100X Secondary Antibody-Peroxidase Conjugate (E) with 5.940 ml of Antibody Diluting Buffer (F) to yield 6 ml of 1X Secondary Antibody-Peroxidase Conjugate. Fifty microliters (50 μl) are needed per well. Allow extra quantity for reservoirs, tubing, pipetting, etc.
- f. **Prepare wash solution:** Prepare 1X Wash Solution by diluting 1 part of the 10X Wash Solution Concentrate (H) with 9 parts of deionized or distilled water. Approximately 1.8 ml are needed per well. Allow extra quantity for reservoirs, tubing, pipetting, etc.

Test Procedure

1. **Load controls and serum samples:** Using a pipettor set at 50 μl , transfer

diluted controls and serum samples into the Antigen-Coated Plate (A). Serum samples and controls should be loaded into the Antigen-Coated Plate (A) as quickly as possible. Change tips between samples. Tap the side of the Antigen-Coated Plate (A) several times to make sure the samples coat the bottom of the wells. Use care not to spill samples from well to well. Incubate the plate 30 minutes at room temperature ($23 \pm 2^\circ\text{C}$).

2. **Wash wells:** After the 30-minute incubation, wash the plate 3 times:
If an automatic washer is used, place the plate on the washing apparatus and wash plate 3 times, filling the wells each time with 1X Wash Solution.
If manual washing is used, dump well contents and remove remaining sera and controls by sharply striking the inverted plate 4 times on a clean paper towel, striking a clean area each time. Immediately fill each well with 1X Wash Solution using a multichannel filling device or a wash bottle. Empty the wash solution from the plate and strike the inverted plate sharply on a clean paper towel as above. Fill and empty the plate by the same method 2 additional times for a total of 3 washes.
3. **Add primary antibody:** Add 50 μl of diluted (1X) Primary Antibody to each well. Tap the side of the loaded assay plate several times to make sure the primary antibody coats the bottom of the wells. Incubate for 30 minutes at room temperature ($23 \pm 2^\circ\text{C}$).
4. **Wash wells:** After the 30-minute incubation, wash the plate 3 times as in Step 2.
5. **Add secondary antibody-peroxidase conjugate:** Add 50 μl of diluted (1X) Secondary Antibody-Peroxidase Conjugate to each well. Tap the side of the loaded assay plate several times to make sure the conjugate coats the bottom of the wells. Incubate for 30 minutes at room temperature ($23 \pm 2^\circ\text{C}$).
6. **Wash wells:** After the 30-minute incubation, wash the plate 3 times as in Step 2.
7. **Add substrate solution:** Add 50 μl of Substrate Solution (I) to each well. Tap the side of the loaded assay plate several times to make sure the substrate coats the bottom of the wells. Incubate 15 minutes at room temperature ($23 \pm 2^\circ\text{C}$). Avoid leaving the plate in direct sunlight. *Do not empty wells.*
8. **Add stop solution:** Add 50 μl of Stop Solution (J) to each well. Tap the side of the loaded assay plate several times to mix the Substrate Solution and the Stop Solution. *Do not empty wells.*
9. **Read and record the test results:** Immediately after adding the Stop Solution, the plate should be read on a microplate absorbance spectrophotometer. Set the optical density (OD) reading wavelength to 620, 630 or 650 nm and read plate(s). Some readers require an empty well on the plate for blanking. In this case, no reagents should be added to this well.
10. Return all remaining kit reagents to 2-7°C for storage.

Calculation of % Inhibition (%I):

$$\% \text{ Inhibition (\%I)} = 100 [1 - (\text{Sample OD} \div \text{NC OD})]$$