

Overview of Standard protocol



Sample types: blood, plasma, serum, milk, oral fluids, ear notch in pigs, tissue homogenized in PBS

1. Sample Lysis

- ✓ 200 µL sample
- ✓ 180 µL SLS
- ✓ 20 µL Proteinase K
- ✓ 4 µL carrier RNA (recommended)
- ✓ x µL Extraction control (recommended)

- 1.1. Mix by pipetting
- 1.2. Add entire lysate to Step 2 Binding Mix

2. Binding of nucleic acids to Beads

- ✓ 600 µL Binding Buffer
- ✓ 20 µL Beads

- 2.1. Mix by shaking for 5-10 min at RT (optional: mix by pipetting)
- 2.2. Remove supernatant after 2 min magnetic separation

3. Washing with Wash Buffer 1

- ✓ 600 µL Wash Buffer 1

- 3.1. Resuspend: Shake for 1-3 min at RT
- 3.2. Remove supernatant after 2 min magnetic separation

4. Washing with Wash Buffer 2

- ✓ 600 µL Wash Buffer 2

- 4.1. Resuspend: Shake for 1-3 min at RT
- 4.2. Remove supernatant after 2 min magnetic separation

5. Washing with 80% Ethanol

- ✓ 600 µL 80% Ethanol

- 5.1. Resuspend: Shake for 1-3 min at RT
- 5.2. Remove supernatant after 2 min magnetic separation

6. Air-drying of Beads

- 6.1. Air dry for 10 min at RT

7. Elution of RNA/DNA

- ✓ 50-100 µL Elution Buffer

- 7.1. Resuspend: Shake for 5 min at RT (optional: mix by pipetting)
- 7.2. Transfer RNA/DNA into elution plate or tube after 2 min magnetic separation



Overview of Enhanced protocol



Sample types: blood, plasma, serum, milk, oral fluids, ear notch in pigs, tissue homogenized in PBS

1. Sample Lysis

- ✓ 200 µL sample
 - ✓ 350 µL ELS
 - ✓ 150 µL PBS
 - ✓ 7 µL carrier RNA (recommended)
 - ✓ x µL Extraction control (recommended)
 - ✓ 1 vial Lysis Beads
- 1.1.** Bead beating for 15 min.
 - 1.2.** centrifugation at 16,000 x g for 5 min.
 - 1.3.** Add 405 µL clarified lysate to Step 2 Binding Mix

2. Binding of nucleic acids to Beads

- ✓ 600 µL Binding Buffer
 - ✓ 20 µL Proteinase K
 - ✓ 20 µL Beads
- 2.1.** Mix by shaking for 5-10 min at RT (optional: mix by pipetting)
 - 2.2.** Remove supernatant after 2 min magnetic separation

3. Washing with Wash Buffer 1

- ✓ 600 µL Wash Buffer 1
- 3.1.** Resuspend: Shake for 1-3 min at RT
 - 3.2.** Remove supernatant after 2 min magnetic separation

4. Washing with Wash Buffer 2

- ✓ 600 µL Wash Buffer 2
- 4.1.** Resuspend: Shake for 1-3 min at RT
 - 4.2.** Remove supernatant after 2 min magnetic separation

5. Washing with 80% Ethanol

- ✓ 600 µL 80% Ethanol
- 5.1.** Resuspend: Shake for 1-3 min at RT
 - 5.2.** Remove supernatant after 2 min magnetic separation

6. Air-drying of Beads

- 6.1.** Air dry for 10 min at RT

7. Elution of RNA/DNA

- ✓ 50-100 µL Elution Buffer
- 7.1.** Resuspend: Shake for 5 min at RT (optional: mix by pipetting)
 - 7.2.** Transfer RNA/DNA into elution plate or tube after 2 min magnetic separation



Overview of Modified protocol



Sample types: ear notch in SLS, tissue homogenized in SLS, ear notch in ELS, tissue homogenized in ELS

1. Sample Lysis

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| <ul style="list-style-type: none">✓ Tissue section✓ 200 µL SLS or ELS✓ 4 µL carrier RNA | <ul style="list-style-type: none">1.1. Incubation for <u>minimum 15 min</u>1.2. Add entire lysate to Step 2 Binding Mix |
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2. Binding of nucleic acids to Beads

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| <ul style="list-style-type: none">✓ 600 µL Binding Buffer✓ 20 µL Proteinase K✓ 20 µL Beads✓ x µL extraction control (recommended) | <ul style="list-style-type: none">2.1. Mix by shaking for <u>5-10 min</u> at RT (optional: mix by pipetting)2.2. Remove supernatant <u>after 2 min</u> magnetic separation |
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3. Washing with Wash Buffer 1

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| <ul style="list-style-type: none">✓ 600 µL Wash Buffer 1 | <ul style="list-style-type: none">3.1. Resuspend: Shake for <u>1-3 min</u> at RT3.2. Remove supernatant <u>after 2 min</u> magnetic separation |
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4. Washing with Wash Buffer 2

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| <ul style="list-style-type: none">✓ 600 µL Wash Buffer 2 | <ul style="list-style-type: none">4.1. Resuspend: Shake for <u>1-3 min</u> at RT4.2. Remove supernatant <u>after 2 min</u> magnetic separation |
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5. Washing with 80% Ethanol

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| <ul style="list-style-type: none">✓ 600 µL 80% Ethanol | <ul style="list-style-type: none">5.1. Resuspend: Shake for <u>1-3 min</u> at RT5.2. Remove supernatant <u>after 2 min</u> magnetic separation |
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6. Air-drying of Beads

- 6.1.** Air dry for 10 min at RT

7. Elution of RNA/DNA

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| <ul style="list-style-type: none">✓ 50-100 µL Elution Buffer | <ul style="list-style-type: none">7.1. Resuspend: Shake for <u>5 min</u> at RT (optional: mix by pipetting)7.2. Transfer RNA/DNA into elution plate or tube <u>after 2 min</u> magnetic separation |
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Overview of High Inhibitors protocol



Sample types: feces

1. Sample Lysis

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| <ul style="list-style-type: none">✓ 140 µL of clarified feces✓ 350 µL Enhanced Lysis Solution✓ 210 µL 1X PBS✓ 7 µL Carrier RNA (recommended)✓ x µL extraction control (recomm.)✓ 1 vial Lysis Beads | <ul style="list-style-type: none">1.1. Bead-beating for 15 min1.2. Centrifugation at 16,000 x g <u>for 5 min</u>1.3. Add 405 µL clarified lysate to Step 2 Binding Mix |
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2. Binding of nucleic acids to Beads

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| <ul style="list-style-type: none">✓ 600 µL Binding Buffer✓ 20 µL Beads✓ 20 µL Proteinase K | <ul style="list-style-type: none">2.1. Mix by shaking <u>for 5-10 min</u> at RT (optional: mix by pipetting)2.2. Remove supernatant <u>after 2 min</u> magnetic separation |
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3. Washing with Wash Buffer 1

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| <ul style="list-style-type: none">✓ 600 µL Wash Buffer 1 | <ul style="list-style-type: none">3.1. Resuspend: Shake <u>for 1-3 min</u> at RT3.2. Remove supernatant <u>after 2 min</u> magnetic separation |
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4. Washing with Wash Buffer 2

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| <ul style="list-style-type: none">✓ 600 µL Wash Buffer 2 | <ul style="list-style-type: none">4.1. Resuspend: Shake <u>for 1-3 min</u> at RT4.2. Remove supernatant <u>after 2 min</u> magnetic separation |
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5. Washing with 80% Ethanol

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| <ul style="list-style-type: none">✓ 600 µL 80% Ethanol | <ul style="list-style-type: none">5.1. Resuspend: Shake <u>for 1-3 min</u> at RT5.2. Remove supernatant <u>after 2 min</u> magnetic separation |
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6. Air-drying of Beads

- 6.1.** Air dry for 10 min at RT

7. Elution of RNA/DNA

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| <ul style="list-style-type: none">✓ 50-100 µL Elution Buffer | <ul style="list-style-type: none">7.1. Resuspend: Shake <u>for 5 min</u> at RT (optional: mix by pipetting)7.2. Transfer RNA/DNA into elution plate or tube <u>after 2 min</u> magnetic separation |
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Overview of High Inhibitors Viral protocol



Sample types: feces

1. Sample Lysis

- ✓ 140 µL of clarified feces
- ✓ 350 µL Enhanced Lysis Solution
- ✓ 7 µL Carrier RNA (recommended)
- ✓ x µL extraction control (recomm.)
- ✓ 1 vial Lysis Beads

1.1. Mix vigorously by pipetting or vortexing

1.2. Centrifugation at 16,000 x g for 5 min

1.3. Add 405 µL clarified lysate to Step 2 Binding Mix

2. Binding of nucleic acids to Beads

- ✓ 600 µL Binding Buffer
- ✓ 20 µL Beads
- ✓ 20 µL Proteinase K

2.1. Mix by shaking for 5-10 min at RT (optional: mix by pipetting)

2.2. Remove supernatant after 2 min magnetic separation

3. Washing with Wash Buffer 1

- ✓ 600 µL Wash Buffer 1

3.1. Resuspend: Shake for 1-3 min at RT

3.2. Remove supernatant after 2 min magnetic separation

4. Washing with Wash Buffer 2

- ✓ 600 µL Wash Buffer 2

4.1. Resuspend: Shake for 1-3 min at RT

4.2. Remove supernatant after 2 min magnetic separation

5. Washing with 80% Ethanol

- ✓ 600 µL 80% Ethanol

5.1. Resuspend: Shake for 1-3 min at RT

5.2. Remove supernatant after 2 min magnetic separation

6. Air-drying of Beads

6.1. Air dry for 10 min at RT

7. Elution of RNA/DNA

- ✓ 50-100 µL Elution Buffer

7.1. Resuspend: Shake for 5 min at RT (optional: mix by pipetting)

7.2. Transfer RNA/DNA into elution plate or tube after 2 min magnetic separation



Overview of High Extenders protocol



Sample types: semen

1. Sample Lysis

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| <ul style="list-style-type: none">✓ 200 µL semen sample✓ 200 µL ELS✓ 20 µL Proteinase K✓ 4 µL carrier RNA (recommended) | <ul style="list-style-type: none">1.1. Incubate at RT for 15 min (optional: Incubate at 60°C <u>for 15 min</u>)1.2. Centrifugation at 15,000 x g <u>for 1 min</u>1.3. Add entire lysate to Step 2 Binding Mix |
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2. Binding of nucleic acids to Beads

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| <ul style="list-style-type: none">✓ 600 µL Binding Buffer✓ 20 µL Beads✓ x µL extraction control (recommended) | <ul style="list-style-type: none">2.1. Mix by shaking <u>for 5-10 min</u> at RT (optional: mix by pipetting)2.2. Remove supernatant <u>after 2 min</u> magnetic separation |
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3. Washing with Wash Buffer 1

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| <ul style="list-style-type: none">✓ 600 µL Wash Buffer 1 | <ul style="list-style-type: none">3.1. Resuspend: Shake <u>for 1 min</u> at RT3.2. Remove supernatant <u>after 2 min</u> magnetic separation |
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4. Washing with Wash Buffer 2

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| <ul style="list-style-type: none">✓ 600 µL Wash Buffer 2 | <ul style="list-style-type: none">4.1. Resuspend: Shake <u>for 1 min</u> at RT4.2. Remove supernatant <u>after 2 min</u> magnetic separation |
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5. Washing with 80% Ethanol

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| <ul style="list-style-type: none">✓ 600 µL 80% Ethanol | <ul style="list-style-type: none">5.1. Resuspend: Shake <u>for 1 min</u> at RT5.2. Remove supernatant <u>after 2 min</u> magnetic separation |
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6. Air-drying of Beads

- 6.1.** Air dry for 10 min at RT

7. Elution of RNA/DNA

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| <ul style="list-style-type: none">✓ 50-100 µL Elution Buffer | <ul style="list-style-type: none">7.1. Resuspend: Shake <u>for 5 min</u> at RT (optional: mix by pipetting)7.2. Transfer RNA/DNA into elution plate or tube <u>after 2 min</u> magnetic separation |
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